Genetic history of cystic fibrosis mutations in Italy. I. Regional distribution

S. RENDINE¹, F. CALAFELL¹, N. CAPPELLO¹, R. GAGLIARDINI², G. CARAMIA², N. RIGILLO³, M. SILVETTI⁴, M. ZANDA⁴, A. MIANO⁵, F. BATTISTINI⁵, L. MARIANELLI⁶, G. TACCETTI⁶, M. C. DIANA⁷, L. ROMANO⁷, C. ROMANO⁷, A. GIUNTA⁸, R. PADOAN⁸, A. PIANAROLI⁸, V. RAIA⁹, G. DE RITIS⁹, A. BATTISTINI¹⁰, G. GRZINCICH¹⁰, L. JAPICHINO¹¹, F. PARDO¹¹, M. ANTONELLI¹², S. QUATTRUCCI¹², V. LUCIDI¹³, M. CASTRO¹³, B. SANTINI¹⁴, M. CASTELLO¹⁴, G. GUANTI¹⁵, G. B. LEONI¹⁶, A. CAO¹⁶, C. TOFFOLI¹⁷, E. LUCCI¹⁷, C. VULLO¹⁷, F. TORRICELLI¹⁸, F. SBERNINI¹⁸, G. ROMEO¹⁹, P. RONCHETTO¹⁹, M. SEIA²⁰, A. ROSSI²⁰, M. FERRARI²¹, L. CREMONESI²¹, F. SALVATORE²², G. CASTALDO²², E. D'ALCAMO²³, A. MAGGIO²³, F. SANGIUOLO²⁴, B. DALLAPICCOLA²⁴, P. MACERATESI²⁴, L. BISCEGLIA²⁵, P. GASPARINI²⁵, A. CARBONARA¹, A. BONIZZATO²⁶, G. CABRINI²⁶, C. BOMBIERI²⁷, P. F. PIGNATTI²⁷, G. BORGO²⁸, C. CASTELLANI²⁸, A. VILLANI²⁸, C. ARDUINO²⁹, D. SALVATORE³⁰, G. MASTELLA²⁸ AND A. PIAZZA^{1,*}

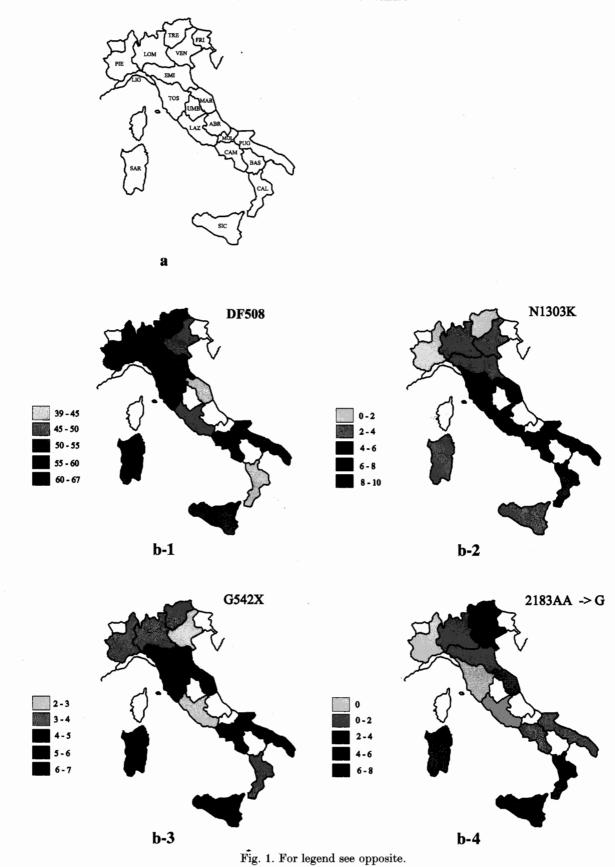
¹Dipartimento di Genetica, Biologia e Biochimica and Centro CNR-CIOS, Torino; ²Ospedale Dei Bambini 'G. Salesi', Ancona; ³ Clinica Pediatrica, Università, Bari; ⁴ Div. Pediatria, Ospedale, Cagliari; ⁵ Centro Fibrosi Cistica, Osp. 'M. Bufalini', Cesena; ⁶ Ospedale 'Mayer', Firenze; ⁷ Clinica Pediatrica Istituto 'G. Gaslini', Genova; 8 Centro Regionale Fibrosi Cistica, IIº Clinica Pediatrica, Università, Milano; ⁹ Dipartimento Pediatria, Università Federico II, Napoli; ¹⁰ Clinica Pediatrica, Università, Parma; 11 Ospedale Dei Bambini 'Di Cristina', Palermo; 12 Centro Fibrosi Cistica, Regione Lazio, Ist. Clinica Pediatrica, Università 'La Sapienza', Roma; 18 Ospedale 'Bambin Gesù', Roma; 14 Ospedale 'Regina Margherita', Clinica Pediatrica, Università, Torino; 15 Ist. Genetica Medica, Università, Bari; 16 Ist. Clinica e Biologia Età Evolutiva, Università, Cagliari; 17 Div. Pediatrica, Arcispedale 'S. Anna', Ferrara; 18 U.O. Citogenetica e Genetica, Azienda Ospedale Careggi, Firenze; 19 Lab. Genetica Molecolare, Ist. 'G. Gaslini', Genova; 20 Laboratorio, Istituti Clinici Perfezionamento, Università, Milano; ²¹I.R.C.C.S. Ospedale San Raffaele, Lab. Biologia Molecolare Clinica, Milano; ²² G.E.I.N.G.E., Biotecnologie Avanzate e Dip. Biochimica e Biotecnologie Mediche, Università Federico II, Napoli; ²³ Serv. Talassemie, Ospedale 'V. Cervello', Palermo; ²⁴ Dip. Sanità Pubblica e Biologia Cellulare, Cattedra Genetica Umana, Università 'Tor Vergata', Roma; ²⁵ Serv. Gen. Med. IRCSS 'Casa Sollievo dalla Sofferenza', San Giovanni Rotondo, Foggia; 26 Lab. Genetica Molecolare, Centro Fibrosi Cistica, Osp. Maggiore, Verona; ²⁷ Ist. Biologia e Genetica, Università, Verona; 28 Centro Fibrosi Cistica, Osp. Maggiore, Verona; 29 Servizio Universitario Genetica Medica, Azienda S. Giovanni, Torino; 30 Serv. Pediatria, Osp. Villa D'Agri, Potenza

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SUMMARY

Earlier analysis of the Italian population showed patterns of genetic differentiation that were interpreted as being the result of population settlements going back to pre-Roman times. DNA disease mutations may be a powerful tool in further testing this hypothesis since the analysis of diseased individuals can detect variants too rare to be resolved in normal individuals. We present data on the relative frequencies of 60 cystic fibrosis (CF) mutations in Italy and the geographical distribution of the 12 most frequent CF mutations screened in 3492 CF chromosomes originating in 13 Italian regions. The 12 most frequent mutations characterize about 73% of the Italian CF chromosomes. The most common mutation, Δ F508, has an average frequency of 51%, followed by

^{*} Correspondence: Prof. Alberto Piazza, Dipartimento di Genetica, Biologia e Biochimica, Via Santena 19, 10126. Torino, Italy. Tel. +39-11-6706650; Fax +39-11-674040. E-mail: Piazza@cios.to.cnr.it



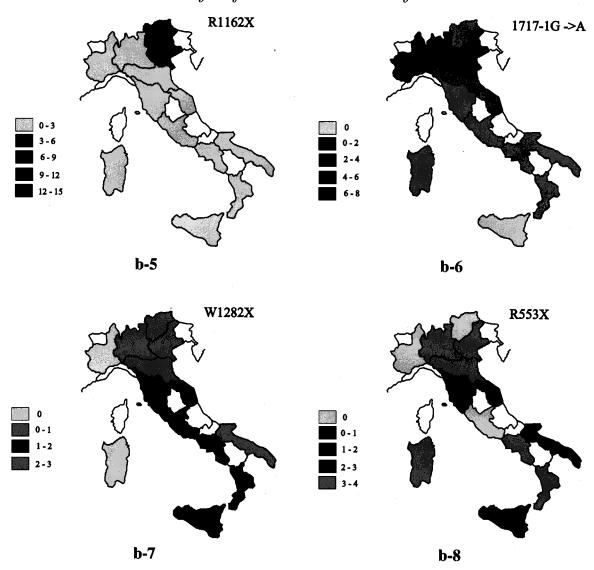


Fig. 1. a, Italian regions: ABR, Abruzzo; BAS, Basilicata; CAL, Calabria; CAM, Campania; EMI, Emilia Romagna; FRI, Friuli; LAZ, Lazio; LIG, Liguria; LOM, Lombardia; MAR, Marche; MOL, Molise; PIE, Piemonte; PUG, Puglia; SAR, Sardegna (Sardinia Island); SIC, Sicilia; TOS, Toscana; TRE, Trentino Alto Adige; UMB, Umbria; VEN, Veneto. b-1 to b-8, Geographical distribution of the most common CF mutations in Italy. Regions with less than 100 chromosomes tested were left blank for their high sampling errors.

N1303K and G542X, both with average frequencies around 5%. Multivariate analyses show that the relative frequencies of CF mutations are heterogeneous among Italian regions, and that this heterogeneity is weakly correlated with the geographical pattern of non-DNA 'classical' genetic markers. The northern regions are well differentiated from the central-southern regions and within the former group the western and eastern regions are remarkably distinct. Moreover, Sardinia shows the presence of mutation T338I, which seems absent in any other European CF chromosome. The north-western regions of Italy, characterized by the mutation $1717-1G \rightarrow A$, were under Celtic influence, while the north-east regions, characterized by the mutations R1162X, 2183AA \rightarrow G and $711+5G \rightarrow A$, were under the influence of the Venetic culture.

INTRODUCTION

An analysis of genetic data may be helpful in tracing historical and prehistorical events (Nei & Roychoudhury, 1982; Cavalli-Sforza et al. 1988, 1993, 1994). An earlier genetic study of the Italian population, based on blood group and protein polymorphisms, showed patterns of differentiation that were interpreted as being the result of events dating back to pre-Roman times (Piazza et al. 1988). Expressed markers, however, may reflect a small fraction of the DNA variation and DNA disease mutations may be a powerful tool to dissect it: the analysis of patients can detect variants too rare to be resolved in samples of non-affected individuals (Dianzani et al. 1994). We have studied the geographical distribution of several cystic fibrosis (CF; MIM 219700) mutations in Italy. CF is the most common autosomal recessive lethal disorder in Caucasoid populations, with an average incidence of about 1 in 2500 individuals (Welsh et al. 1995) and a carrier rate of 1 in 25. CF is caused by mutations in a 27 exon gene spanning about 250 kb on chromosome 7 (region q31.3) and codes for a 1480 amino acid protein named Cystic Fibrosis Transmembrane conductance Regulator (CFTR) (Riordan et al. 1989). To date more than 600 CF mutations have been described in the world population, among which $\Delta F508$ is the most common, with an average relative frequency among Caucasoids of about 66% (Cystic Fibrosis Genetic Analysis Consortium, 1994) http://genet.sickkids.on.ca). Most mutations are rare and either have specific geographical or ethnical patterns or are sporadic. The $\Delta F508$ mutation is present in about 80% of North European CF chromosomes (87% in Denmark) and only in around 50% of South European chromosomes (European Working Group on CF Genetics, 1990; Gasparini et al. 1990; Cystic Fibrosis Genetic Analysis Consortium, 1994). This means that other mutations are expected to be found in Mediterranean populations.

The aim of this work is to present an analysis of CF mutation relative frequencies in the Italian regions. Our results might prove of practical use when designing strategies for DNA analysis of

patients for diagnostic purpose, for prenatal diagnosis of CF cases, and when planning population screening for heterozygotes: the carrier status cannot be detected clinically or biochemically, but only by mutation analysis. Therefore a screening programme (if considered) should be planned on the basis of the regional mutation prevalence. Our sampling, however, may be biased in regions with few affected individuals and, in such regions, the possible order of mutations to be tested must be considered with some caution.

A less practically oriented but nevertheless interesting hypothesis to test is whether the geographical patterns of the mutations causing this genetic disorder retrace those shown by non-DNA-polymorphisms: the history of a single gene and the genetic history of a population do not necessarily follow the same pattern, as the date of a single mutation does not provide, by itself, a reference to date the separation of human demic units.

The present report resulted from a joint collaboration of 14 Italian laboratories and 15 CF centres, and adds new data to those already published (Cremonesi et al. 1990; Nunes et al. 1991; Gasparini et al. 1993; Bonizzato et al. 1995) and completes the picture of CF mutation distribution in Italy. This is, to the best of our knowledge, the second largest CF mutation data set for a single country after that published by Schwarz et al. (1995).

MATERIALS AND METHODS

Subjects

A sample of 1746 CF patients was collected. The geographic origin of all the 3492 CF chromosomes was established by assessing the birthplace of the four grandparents assigning each chromosome (or fraction of it, if the grandparents giving origin to it were born in different places) to one of the Italian regions defined in the first column of Table 3 and represented in Figure 1a. When families included more than one CF patient, only one of them was randomly selected. No patients from consanguineous marriages were included.

Amplification refractory mutation	system									Yes				(Ferrie $et\ al.$ 1992)	
Restriction enzyme	analysis					Λ es		Λ es	m Aes	m Yes	Λ es	m Yes		(Haliassos $et al. 1989$)	•
Denaturing gel gradient	electrophoresis				Yes	Yes			Yes	Yes	m Yes		m Yes	(Fanen $et al.$ 1992)	•
Restriction site generating				Yes			m Yes						m Yes	(Gasparini et al. 1992)	
DNA	immunoassay	Yes	Yes	Yes			Yes							(Sangiuolo et al. 1995)	
Allele specific oligonucleotide dot_blot						Yes	Yes	Yes	Yes	Yes	Yes	Yes	m Yes	(Castaldo $et al. 1996$)	
Reverse	dot blot	Yes	Yes	Yes			Yes	Yes	Yes	Yes				(Saiki <i>et</i> <i>al.</i> 1989)	
DNA heteroduplex	analysis	Yes												(Rommens $et al. 1990$)	
	Mutation	Δ F508	N1303K	G542X	$2183AA \rightarrow G$	R1162X	$1717-1G \rightarrow A$	W1282X	R553X	T338I	R347P	$711 + 5G \rightarrow A$	$621+1G \rightarrow T$	Reference	

Mutation analysis

Sixty different CF mutations were screened. Their choice depended on criteria adopted by the single laboratories on the basis of their technical experience and of frequency data reported either locally or in other European populations.

In a preliminary data exploration, it became apparent that most mutations were restricted to one or few chromosomes. We consider for further analysis only those 12 mutations that had been found in at least 1% of the chromosomes in one or more regions.

Table 1 shows the methods of CF mutation detection adopted by the laboratories to identify the 12 most frequent mutations.

As collection of data was partially retrospective, the set of mutations tested in the laboratories was not exactly the same. Those individuals in which two mutations were found were presumed to be negative for all other mutations. The relative frequency of the mutation which was not tested in all laboratories, was then calculated by the ratio M/N, where M is the number of the chromosomes carrying that mutation and N is the sum of two numbers: (a) the number of chromosomes which were actually tested; and (b) the number of chromosomes which, even if not actually tested, almost certainly do not carry that mutation as the individuals were found positive for two other mutations. We have lost the information from the laboratories which did not test this mutation and provided cases of CF patients with at least one chromosome negative for all the mutations they actually tested. This is the reason why the mutation frequencies are calculated from numbers of chromosomes which varied from a minimum of 1824 to a maximum of 3442, depending on the mutation. We are well aware that this method of calculating mutation frequencies gives biased results in the sense of systematically underestimating the less frequent mutations, but simulation experiments (not shown) indicate that such a bias does not substantially affect our results.

Methods of genetic analysis

Only the 13 regions with sample sizes higher than 100 chromosomes (Table 3) were included in multivariate analyses based on the 12 more frequent CF mutations.

- Principal component analysis (Hotelling, 1933) was used to reduce the dimensional space of the 12 mutations into a twodimensional space with the least distortion so that the relative position of the Italian regions in this space gives the best possible representation of their mutual differences.
- 2. Dendograms are very popular ways of displaying and clustering objects (in our case the Italian regions) under the form of trees of descent even if no phylogenetic relationship holds. among them There algorithms to reconstruct trees, depending on which model of evolution among the objects is assumed. We used the methods of neighbourjoining (Saitou & Nei, 1987), based on Reynolds et al.'s (1983) genetic distances, and maximum likelihood (Felsenstein, 1981). The maximum likelihood method to reconstruct trees is by definition the best when allele frequencies are used and in fact it has been applied to the frequencies of the haplotypes involving the CF gene region microsatellites (Estivill et al. 1994) but its optimal behaviour is doubtful when relative frequencies of mutations are used.
- 3. The robustness of a dendrogram representation was tested by the bootstrap technique (Felsenstein, 1985; Efron & Tibshirani, 1993). The relative frequency of each splitting into 500 bootstrapped tree representations was calculated and indicated in the displayed dendrogram, a percentage larger than 50% indicating a reliable clustering.

Genetic distances and dendograms were produced by using the PHYLIP 3.5c package (Felsenstein, 1989).

RESULTS

The 60 different mutations screened in our Italian sample of CF patients are shown in Table

2 with their frequencies. We estimated (last column of table 2) that about 77% of CF chromosomes can be attributed to these 60 mutations. Among these, 20 mutations were found in 2-10 chromosomes and 28 mutations are present only in one chromosome. Note, however, a possible source of bias, specially in the range of low frequencies, the number of cases negative for one mutation being somewhat correlated with the number of positive cases ascertained for another mutation, as CF patients with two mutations identified have been assigned negative for all mutations even though they were not actually tested for those mutations. Table 3 shows the distribution in 19 Italian regions of the relative frequencies of the 12 more frequent CF mutations: Δ F508, N1303K, G542X, 2183AA \rightarrow G, R1162X, 1717-1G \rightarrow A, W1282X, R553X, T338I, R347P, $711 + 5G \rightarrow A$, $621 + 1G \rightarrow T$. No CF chromosomes with origin in Valle d'Aosta region were found.

The most common mutation in Italy is $\Delta F508$, with an average frequency of 51%. The highest frequencies are found in Piemonte, Umbria and Campania (>60%), the lowest in Friuli and Marche (<40%). The next more frequent mutations are N1303K and G542X, with mean frequencies of 4.84% and 4.83% respectively. The N1303K mutation is relatively common in central and south-eastern regions (Abruzzo, Marche, Lazio, Molise and Puglia), frequencies between 8% and 10%; it is rare in Piemonte and Trentino Alto Adige (<1%). G542X shows frequencies ranging from 2% (Trentino Alto Adige) to 13%; frequencies higher than 10% are found in three regions (Basilicata, Friuli and Liguria) with a low number of chromosomes tested. The 2183AA → G mutation, with a mean frequency of 3%, is the fourth most common mutation in Italy, reaching its highest frequencies (7%) in Trentino Alto Adige and Veneto. Mutation R1162X shows an average frequency of 2.42%, but reaches frequencies of 14% in Trentino Alto Adige and 9% in Veneto. The 1717-1G \rightarrow A mutation accounts for 4–6% of the CF chromosomes in Lombardia, Piemonte and Liguria, but is rare or absent in the central and southern regions (< 2%).

Table 2. Frequencies of the 60 different mutations screened in the Italian sample of CF patients

			1 , V
Mutation	Number of chromosomes tested	Frequency	Cumulative frequency
ΔF508	3442	0·5107	- •
N1303K	3056	0.0484	$0.5107 \\ 0.5591$
G542X	3082	0.0483	0.6075
2183 AA-> G	2596	0.0266	0.6341
R1162X	$\begin{array}{c} 2350 \\ 2850 \end{array}$	0.0242	0.6583
1717-1G-> A	$\begin{array}{c} 2892 \\ \end{array}$	0.0242	0.6794
W1282X	2600	0.0123	0.6917
R553X	2882	0.0115	0.7031
T338I	2306	0.0069	0.7101
R347P	2642	0.0061	0.7161
711+5G-> A	2454	0.0057	0.7218
G85E	1980	0.0040	0.7259
621 + 1G -> T	2594	0.0039	0.7297
R334W	2366	0.0030	0.7327
R352Q	2112	0.0024	0.7350
S549N	2118	0.0024	0.7374
R347H	2184	0.0018	0.7392
L1077P	1840	0.0016	0.7409
R1158X	1878	0.0016	0.7425
$541 \mathrm{delC}$	1884	0.0016	0.7440
R1066H	1918	0.0016	0.7456
E585X	1922	0.0016	0.7472
Q552X	2172	0.0014	0.7486
D1152H	1824	0.0011	0.7497
2790-2A->G	1862	0.0011	0.7507
3132 del TG	1862	0.0011	0.7518
3667ins4	1876	0.0011	0.7529
DelI507	1914	0.0010	0.7539
1898 + 3A -> G	1920	0.0010	0.7550
G1244E	1960	0.0010	0.7560
1784delG	2052	0.0010	0.7569
G551D	2600	0.0008	0.7577
4382delA	1822	0.0005	0.7583
$2184 \mathrm{insG} \ \mathrm{H}139 \mathrm{R}$	1822 1822	0.0005	0.7588
711+3AG	1822 1824	$0.0005 \\ 0.0005$	$0.7594 \\ 0.7599$
L558S	1836	0.0005	0.7605
L1065P	1838	0.0005	0.7603
M348K	1838	0.0005	0.7615
S912X	1854	0.0005	0.7621
1706del17	1856	0.0005	0.7626
G1349D	1856	0.0005	0.7632
F693L	1862	0.0005	0.7637
M1V	1862	0.0005	0.7642
R709X	1862	0.0005	0.7648
1717-8G-> A	1876	0.0005	0.7653
406-1G-> C	1876	0.0005	0.7658
457TAT-> C	1876	0.0005	0.7664
G178R	1876	0.0005	0.7669
D579G	1880	0.0005	0.7674
3849 + 4A -> G	1882	0.0005	0.7680
W57G	1884	0.0005	0.7685
D1270N	1896	0.0005	0.7690
3849 + 10 KbC - > T	1912	0.0005	0.7695
I148T	1916	0.0005	0.7701
R1066C	1920	0.0005	0.7706
C524X	1948	0.0005	0.7711
$2909 ext{delT}$	1956	0.0005	0.7716
S549RA -> C	2018	0.0005	0.7721
$1078 \mathrm{delT}$	2136	0.0005	0.7726

Table 3. Relative frequencies and sample sizes of the 12 more frequent CF mutations in the Italian regions

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	$\Delta F508$	N1303K	G542X	2183AA -> G	R1162X	1717-1G-> A	W1282X	R553X	T338I	R347P	711 + 5G -> A	621 + 1G -> T	$\mathbf{Total^f}$
$Abruzzo^c$	64 ^a (49) ^b	58 (10)	51 (6)	38 (0)	50(2)	42 (2)	37 (7)	50 (0)	37 (0)	39 (0)	40 (0)	43 (0)	76
Basilicata ^c	76 (47)	67 (6)	69 (11)	60 (5)	54 (0)	64 (0)	59 (3)	64(0)	45 (0)	54 (0)	61 (0)	66 (0)	72
Calabria	148 (45)	133 (5)	137 (3)	97 (2)	119 (0)	123 (2)	102 (2)	121(1)	102 (0)	112 (3)	97 (0)	115 (0)	63
Campania	223 (61)	216 (7)	215 (4)	182 (2)	197 (0)	207 (1)	182(2)	204(1)	149 (0)	195(0)	193 (0)	200 (0)	78
EmilRom.	241 (54)	214 (3)	226 (6)	168 (1)	198 (1)	212 (2)	176 (1)	212(1)	162 (0)	172(1)	166 (0)	168 (0)	. 69
Friulic •	24 (35)	18 (4)	22 (13)	17 (3)	19 (5)	19 (0)	18 (6)	21 (0)	15 (0)	16 (0)	16 (0)	17 (0)	66
Lazio	235 (46)	225 (10)	219 (3)	164 (0)	178 (0)	184 (1)	127(2)	179 (0)	134 (0)	174 (0)	178 (0)	209 (2)	64
$\mathbf{Liguria^c}$	46 (60)	37 (4)	37 (12)	33 (0)	35 (0)	37 (4)	34 (0)	38 (4)	35 (0)	37 (0)	35 (0)	36 (0)	85
Lombardia	401 (50)	384 (4)	385 (4)	268 (1)	310(1)	385 (6)	362(1)	374(1)	329(0)	350(0)	263 (0)	292 (0)	67
\mathbf{Marche}	144 (39)	130 (9)	135 (6)	113 (2)	116(0)	126 (2)	119 (2)	129(2)	112 (0)	110 (1)	111 (1)	114 (1)	65
$Molise^{c}$	27 (47)	23 (9)	24 (4)	19 (0)	20 (0)	21 (0)	13 (0)	22(0)	16 (0)	20 (5)	20 (0)	21 (0)	65
Piemonte	117 (66)	109 (0)	108 (4)	97 (0)	106 (0)	104 (5)	104 (0)	108 (0)	76 (0)	108 (0)	99 (0)	102 (0)	75
Puglia	240 (54)	227 (8)	221 (6)	172 (1)	201 (0)	206 (1)	191 (0)	202(2)	161 (0)	183 (0)	171 (0)	184 (0)	72
Sardegna	139 (58)	136 (3)	138 (6)	123 (2)	132 (0)	136 (1)	128 (0)	134 (0)	128 (11)	133 (0)	124 (0)	126 (1)	84
Sicilia	380 (52)	331 (3)	339 (7)	272(2)	301 (0)	330 (0)	318 (3)	333(3)	269(0)	296(2)	274 (0)	282 (0)	73
Toscana	191 (51)	148 (5)	154 (5)	111 (0)	119 (0)	145 (1)	124(2)	145 (2)	96 (0)	121(2)	115 (0)	118 (0)	69
Trentino	113 (51)	102 (1)	101 (3)	106 (7)	103 (14)	98 (1)	88 (1)	93 (0)	83 (0)	90 (0)	95 (6)	89 (0)	85
$\mathbf{Umbria^c}$	37 (67)	35 (3)	35 (10)	29 (0)	29 (0)	31 (0)	29 (0)	32 (3)	27 (0)	30 (0)	28 (0)	30 (0)	83
Veneto	552 (45)	426 (4)	429 (2)	496 (7)	528 (9)	387 (3)	357 (0)	384 (0)	307 (0)	371 (0)	340 (2)	351 (0)	73
$Mixed^d$	44 (61)	37 (4)	37 (4)	31 (0)	35 (1)	35 (0)	32 (0)	37 (3)	23 (0)	31 (0)	28 (1)	31 (0)	74
Italy	3442 (51)	3056 (5)	3082 (5)	2596 (3)	2850(2)	2892 (2)	2600 (1)	2882 (1)	2306 (1)	2642 (1)	2454 (1)	2594 (0)	73
$Italy^e$	3398 (51)	3019 (5)	3045 (5)	2565 (3)	2815 (2)	2857 (2)	2568 (1)	2845 (1)	2283 (1)	2611 (1)	2426 (1)	2563 (0)	73

^a Number of chromosomes tested; ^b percentage; ^c number of chromosomes tested less than 50 for at least one mutation; ^d individuals with at least one grandparent born outside Italy; ^e 'mixed' individuals are not considered; ^f the total number of chromosomes tested is not given, as not all chromosomes were tested for all mutations.

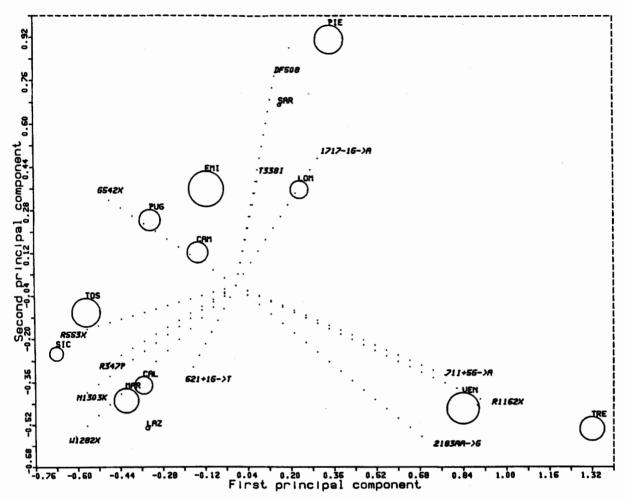


Fig. 2. Biplot of the CF mutation relative frequencies for the 13 Italian regions. Regions and axes of CF mutations (dotted lines) are represented in the space of the first two principal components summarizing 52·1% of the total variation. Each point (region) is at the centre of a circle whose area is inversely proportional to the euclidean distance between the point in this two-dimensional space and the same point in the multidimensional space. The projection of a point (centre of the circle) on a mutation frequency axis gives the relative contribution of that mutation in determining the position of the point in the plot (for more details see Gabriel, 1981).

The six remaining mutations are less frequent (average frequencies near 1%) and more locally distributed: W1282X has high frequencies in Abruzzo and Friuli (both regions have small sample sizes); $711+5G\rightarrow A$ is present only in Marche, Trentino Alto Adige and Veneto; whereas T338I is an exclusively Sardinian mutation. The 48 mutations not selected for further analysis account for $2\cdot6\%$ (93/3492) of CF chromosomes.

As mentioned before, most but not all chromosomes were tested for all 12 mutations, therefore our frequencies may represent biased estimates of the relative frequencies of the less frequent mutations. It is therefore reassuring

that the results we obtained agree with those reported for a smaller subsample where all the coding and adjoining non-coding portions of the gene were sequenced (Bonizzato *et al.* 1995).

The regional frequencies of the most common mutations are displayed in Figure 1*b*-1 to 1*b*-8. Regions where less than 100 chromosomes were tested were left blank for their high sampling errors.

Figure 2 shows 13 Italian regions in the space of the first two principal components summarizing 52·1% of the original mutation variation with a graphical display of the distortion introduced in the two-dimensional representation. The first principal component, which

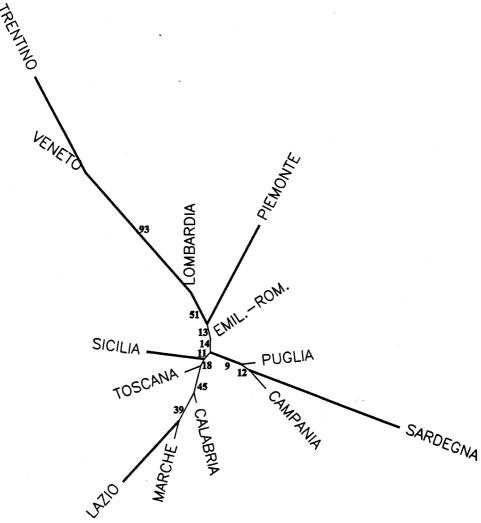


Fig. 3. Dendrogram of 13 Italian regions according to the maximum likelihood method of reconstruction (Felsenstein, 1981). Segments whose lengths are statistically different from zero are drawn with bold lines. Numbers on branches indicate 'bootstrap' percentages, i.e. the relative frequencies of the clusterings in the dendrogram shared among 500 replicates. Percentages higher than 50 reflect a robust structure.

explains 30.68% of the total variation, clearly separates Trentino Alto Adige and Veneto from the other regions. These two regions show high frequencies of three mutations: R1162X, $2183AA \rightarrow G$, and $711+5G \rightarrow A$. The second principal component explains 21.42% of the variance and is highly correlated with the $\Delta F508$ mutation (r=80%); north-western regions of Italy (Piemonte and Lombardia) and Sardinia are differentiated from central and southern regions. The third principal component (16.16% of the variation, not shown) discriminates Lazio, with a relatively high frequency of $621+1G\rightarrow T$, and Sardinia, which shows the 'private' mutation T338I, from the other regions.

A neighbour-joining dendrogram based on Reynolds et al.'s (1983) genetic distances (not shown) groups the 13 Italian regions according to their $\Delta F508$ mutation frequencies: the wide variation in $\Delta F508$ frequencies seems to override the contribution of the other mutations.

A more informative dendrogram can be reconstructed by the maximum likelihood method (Fig. 3). As it uses the variance-covariance matrix between regions, it takes into account the mutation relative frequency variability (variance) within each region. Only one cluster of Figure 3 is robust (bootstrap value 51%): it includes the four northern regions (Trentino Alto Adige, Veneto, Lombardia and

Piemonte). Within this group, Trentino Alto Adige and Veneto form a subcluster with the highest bootstrap value (93%). Central and southern regions are poorly structured, as indicated by the low bootstrap frequencies and by the many segments of the dendrogram whose lengths are statistically not different from zero. The segments leading to Sardinia, Lazio and Sicilia are statistically different from zero for the high frequencies of mutations T338I, $621+1G \rightarrow$ T and R553X respectively.

DISCUSSION

A preliminary point to stress is that we are dealing with 'relative' mutation frequencies, and therefore they do not estimate the frequencies of the mutation in the population if the prevalence of the disease varies from region to region: however, the relative frequencies have the advantage of being roughly independent from the prevalence of the disease (thus from selective factors) and it seemed interesting to evaluate how they work as population markers.

The present study characterized the Italian CF chromosomes, and the geographical pattern of 12 relatively frequent mutations in Italy as given by the birthplaces of the grandparents of the CF cases. One can ask whether this way of distributing the chromosomes among the Italian regions is not only of anthropological value but also of practical use in DNA testing which would require data for the current population. Table 3 has been recalculated by considering the samples who currently live in the various regions (data not shown) and we found that only the regions Abruzzo, Basilicata, Calabria, Friuli, and Umbria show some discrepancies: with the exception of Calabria, in these regions the number of chromosomes tested was less than 50 and in fact they have been excluded from further analyses. In Calabria the relative frequency of the mutation Δ F508 changes from 45% of Table 3 to 51% which is reasonable since Calabria is one of the southern regions most affected by emigration over the last two generations.

As it is well known, the most common mutation is $\Delta F508$, with an average frequency of 51%, followed by N1303K and G542X, both with average frequencies around 48%. This pattern is common to other European countries which share relatively low $\Delta F508$ frequencies but high N1303K and G542X frequencies, and where a high number of mutations accounts for a relatively low fraction of CF chromosomes (Claustres et al. 1993; Chillón et al. 1994; Bertranpetit & Calafell, 1996). An analysis of the relationship between our Italian CF sample and those of other European populations will be reported elsewhere.

Our microgeographic analysis of CF mutation relative frequencies in Italy points to a genetic heterogeneity among Italian regions. Northern regions are well differentiated from central-southern regions (Figs 2, 3). Within the former group two subclusters show well defined identities: the north-western regions (Piemonte and Lombardia) and the north-eastern regions (Trentino Alto Adige and Veneto). Sardinia, known to be genetically very different from the continental Italian population and from other European populations (Piazza et al. 1988), shows the presence of the 'private' mutation T338I, which, to the best of our knowledge, has not been identified in any other CF sample.

A relevant question to investigate is whether the frequency of the CF gene is homogeneous among the Italian regions. Very unfortunately we do not have reliable gene frequencies at a regional level which take into account the region of origin of the patients. The gene frequencies inferred from consanguineous marriages (Romeo et al. 1985) show figures from 0 02055 to 0 02473 (depending on the calculation method) which seem non-heterogeneous among regions, but the estimates are indirect and it is difficult to evaluate their standard error.

A previous work based on 'classical' non-DNA polymorphisms (Piazza et al. 1988) showed that the genetic structure of modern Italy may reflect ancient patterns of ethnic differentiation going back to pre-Roman times. A Greek influence was suggested to have shaped the gene pool of

southern Italian regions. Other well defined civilizations at the end of the sixth century BC played a very important role in the history of Italy and were described as possibly related to important components of the Italian genetic structure: Etrurian in Central Italy; Ligurian in North-western Italy; and Osco-Umbrian in central-eastern regions. At that time North Italy culturally and ethnically characterized areas. The western and central regions (corresponding to modern Valle d'Aosta, Piemonte and Lombardia) were under the Celtic influence; in the Central-eastern Alps (Trentino Alto Adige) lived the Rhaeti, and Veneto was the home of the Venetic culture (Pallottino, 1984).

The distribution of CF mutation relative frequencies shows a geographic pattern that seems similar to that obtained by using non-DNA markers only in North Italy. It is suggested that in the northern regions, near the physical barrier of the Alps, genetic drift played a significant role in determining the high frequencies observed for mutation $1717-1G \rightarrow A$ in central-western regions, and for mutations R1162X, $2183AA \rightarrow G$ and $711+5G \rightarrow A$ in eastern regions.

Central and southern Italy show a more blurred pattern. Genetic distances between Italian regions, when measured from 19 loci and 76 classical gene frequencies (Piazza et al. 1989a, b), are not significantly correlated with CF mutation frequencies with and without ΔF508 (data not shown). A plausible interpretation, not to recall the possibility of recurrent mutations, is that while most polymorphisms of the classical markers probably predate the origin of our species and therefore can accumulate a large variability between ethnic groups, it is probable that CF mutations arose at different and relatively recent times, as suggested for $\Delta F508$ (Morral et al. 1994) and G542X and N1303K (Morral et al. 1993). Another more interesting reason for the discrepancy may be a historical one: the history of South Italy, being connected with the Mediterranean populations, is much more complex than the history of the northern part of Italy, too complex to be reached from CF mutations (taken separately or together) if their date of appearance is not available. An alternative and much more trivial reason may simply be that the size of our sample or the geographical distribution of the disease itself do not allow to dissect what is shared and what is not between the two histories, the history of a single gene from the history of a complex population.

A microsatellite haplotype analysis of Δ F508 carrying chromosomes in Europe was performed to estimate the age of this mutation (Morral et al. 1994): the minimum age for the Δ F508 mutation was estimated at 52000 years ago, that is before Italy and Europe were settled by anatomically modern humans. Watterson & Guess (1977) asked the question whether the most frequent allele is also the oldest. If the frequency and the age of a mutation are proportional, it is suggestive that the time of origin of the mutations typical of North Italy (namely, $1717-1G \rightarrow A$, R1162X, $2183AA \rightarrow G$ and $711+5G \rightarrow A$) are compatible with the spread of Celtic and Venetic speaking people: a more rigorous estimate would be of interest when microsatellite polymorphisms will be available for most mutations.

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REFERENCES

Bertranpetit, J. & Calafell, F. (1996). Genetic and geographical variability in cystic fibrosis: evolutionary considerations. In *Variation in the human genome* (ed.), pp. 97–118. Chichester (Ciba Foundation Symposium 197): Wiley.

Bonizzato, A., Bisceglia, L., Marigo, C., Nicolis, E., Bombieri, C., Castellani, C., Borgo, G., Zelante, L., Mastella, G., Cabrini, G., Gasparini, P. & Pignatti, P. F. (1995). Analysis of the complete coding region of the CFTR gene in a cohort of CF patients from North-Eastern Italy: identification of 90 % of the mutations. Hum. Genet. 95, 397-402.

Castaldo, G., Rippa, E., Sebastio, G., Raia, V., Ercolini, P., De Ritis, G., Salvatore, D. &

- SALVATORE, F. (1996). Molecular epidemiology of cystic fibrosis mutations and respective haplotypes in Southern Italy evaluated with an improved semiautomated robotic procedure. J. Med. Genet. (in press).
- CAVALLI-SFORZA, L. L., PIAZZA, A., MENOZZI, P. & MOUNTAIN, J. L. (1988). Reconstruction of human evolution; bringing together genetic, archaeological, and linguistic data. Proc. Natl. Acad. Sci. USA 85, 6002-6006.
- CAVALLI-SFORZA, L. L., MENOZZI, P. & PIAZZA, A. (1993). Demic expansions and human evolution. Science 259, 639-646.
- CAVALLI-SFORZA, L. L., MENOZZI, P. & PIAZZA, A. (1994).

 History and geography of human genes. Princeton, NJ:
 Princeton University Press.
- CHILLÓN, M., CASALS, T., GIMÉNEZ, J., RAMOS, M. D., PALACIO, A., MORRAL, N., ESTIVILL, X. & NUNES, V. (1994). Analysis of the CFTR gene confirms the heterogeneity of the Spanish population: 43 mutations account for only 78% of CF chromosomes. *Hum. Genet.* 93, 447–451.
- CLAUSTRES, M., MAGUELONE, L., DESGEORGES, M., GIANSILY, M., CULARD, J. F., RAZAKATSARA, G., GERRARD, B. & DEMAILLI, J. (1993). Analysis of the 27 exons and flanking regions of the cystic fibrosis gene: 40 different mutations account for 91.2% of the mutant alleles in Southern France. Hum. Molec. Gen. 2, 1209–1213.
- CREMONESI, L., RUOCCO, L., SEIA, M., RUSSO, S., GIUNTA, A., RONCHETTO, P., FENU, L., ROMANO, L., DEVOTO, M., ROMEO, G. & FERRARI, M. (1990). Frequency of the DF508 mutation in a sample of 175 Italian cystic fibrosis patients. *Hum. Genet.* 85, 400–402.
- Cystic Fibrosis Genetic Analysis Consortium (1994). Population variation of common cystic fibrosis mutations. *Hum. Mutat.* 4, 167–177.
- DIANZANI, I., GIANNATASIO, S., DE SANCTIS, L., MARRA, E., PONZONE, A., CAMASCHELLA, C. & PIAZZA, A. (1994). Genetic history of PKU mutations in Italy. Am. J. Hum. Genet. 55, 849–850.
- EFRON, B. & TIBSHIRANI, R. J. (1993). An introduction to the bootstrap. New York: Chapman & Hall.
- ESTIVILL, X., MORRAL, N. & BERTRANPETIT, J. (1994). Reply to Kaplan, N. L., Lewis, P. O. and Weir, B. S. Nature Genetics 8, 216–218.
- EUROPEAN WORKING GROUP ON CF GENETICS (1990). Gradient of distribution in Europe of the major CF mutation and of its associated haplotypes. *Hum. Genet.* 85, 436-441.
- FANEN, P., GHANEM, N., VIDAUD, M., BESMOND, C., MARTIN, J., COSTES, B., PLASSA, F. & GOOSSENS, M. (1992). Molecular characterization of Cystic Fibrosis: 16 novel mutations identified by analysis of the whole Cystic Fibrosis Conductance Transmembrane Regulator (CFTR) coding regions and spile site junctions. Genomics 13, 770-776.
- Felsenstein, J. (1981). Evolutionary trees from gene frequencies and quantitative characters: finding maximum likelihood estimates. *Evol. Biol.* 35, 1229–1242.
- Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. Evolution 39, 783-791.
- Felsenstein, J. (1989). PHYLIP Phylogeny Inference Package (Version 3.2). Cladistics 5, 164–166.

- FERRIE, R. M., SCHWARZ, M. J., ROBERTSON, N. H., et al. (1992). Development, multiplexing, and application of ARMS tests for common mutations in the CFTR gene. Am. J. Hum. Genet. 51, 251–262.
- Gabriel, K. R. (1981). Biplot display of multivariate matrices for inspection of data and diagnosis. In *Interpreting multivariate data* (ed. B. Barnett), pp. 147–173. Chichester: Wiley.
- Gasparini, P., Pignatti, P. F., Novelli, G., Dallapiccola, B., Nunes, V., Casals, T., Estivill, X., Fernandez, E., Balassopoulou, A., Loukopoulos, D., Lavinaa, J., Sinova, L. & Komel, R. (1990). Mutation analysis in cystic fibrosis. N. Eng. J. Med. 323, 62–63.
- Gasparini, P., Bonizzato, A., Dognini, M. & Pignatti, P. F. (1992). Restriction Site Generating Polymerase Chain Reaction (PCR) for the probeless detection of hidden genetic variation: application to the study of some common cystic fibrosis mutations. *Mol. Cell Probes* 6, 1–7.
- GASPARINI, P., MARIGO, C., BISCEGLIA, G., NICOLIS, E., ZELANTE, L., BOMBIERI, C., BORGO, G., PIGNATTI, P. F. & CABRINI, G. (1993). Screening of 62 mutations in a cohort of CF patients from north eastern Italy: their incidence and clinical features of defined genotypes. Hum. Mutat. 2, 389-394.
- HALIASSOS, A., CHOMEL, G., TESSON, L., et al. (1989).
 Modification of enzymatically amplified DNA for the detection of point mutations. Nuc. Ac. Res. 17, 3606.
- HOTELLING, H. (1933). Analysis of a complex of statistical variables into principal components. J. Educ. Psych. 24, 417-441, 498-520.
- Mantel, N. (1967). The detection of disease clustering and a generalized regression approach. *Cancer Res.* 27, 209–220.
- MORRAL, N., NUNES, V., CASALS, T., CHILLÓN, M., GIMÉNEZ, J., BERTRANPETIT, J. & ESTIVILL, X. (1993). Microsatellite haplotypes for cystic fibrosis: mutation frameworks and evolutionary tracers. *Hum. Molec. Gen.* 2, 1015–1022.
- Morral, N., Bertranpetit, J., Estivill, X., Nunes, V., Casals, T., Giménez, J., Reis, A., Varon-Mateeva, R., Macek, M., Kalaydjieva, L., Angelicheva, D., Dancheva, R., Romeo, G., Russo, M. P., Garnerone, S., Restagno, G., Ferrari, M., Magnani, C., Claustres, M., Desgeorges, M., Schwartz, M., Schwarz, M., Dallapiccola, B., Novelli, G., Ferec, C., de Arce, M., Nemeti, M., Kere, J., Anvret, M., Dahl, N. & Kadasi, L. (1994). The origin of the major cystic fibrosis mutation (DeltaF508) in European populations. Nature Genetics 7, 169–175.
- Nei, M. & Roychoudhury, A. K. (1982). Genetic relationship and evolution of human races. *Evol. Biol.* 14, 1-59.
- Nunes, V., Gasparini, P., Novelli, G., Gaona, A., Bonizzato, A., Sangiuolo, F., Balassopoulou, A., Giménez, F. J., Dognini, M., Ravnik-Glavac, M., Cikuli, M., Mokini, V., Komel, R., Dallapiccola, B., Pignatti, P. F., Loukopoulos, D., Casals, T. & Estivill, X. (1991). Analysis of 14 cystic fibrosis mutations in five South European populations. *Hum. Genet.* 87, 737–738.

- Pallottino, M. (1984). Storia della prima Italia. Milano: Rusconi.
- Piazza, A., Cappello, N., Olivetti, E. & Rendine, S. (1988). A genetic history of Italy. *Ann. Hum. Genet.* **52**, 203–213.
- Piazza, A., Olivetti, E., Barbanti, M., Reali, G., Domenici, R., Giari, A., Benciolini, P., Caenazzo, L., Cortivo, P., Bestetti, A., Bonavita, V., Crinò, C., Pascali, V. L., Fiori, A. & Bargagna, M. (1989 a). The distribution of some polymorphisms in Italy. Gene Geography 3, 69–139.
- PIAZZA, A., OLIVETTI, E., GRIFFO, R. M., RENDINE, S., AMOROSO, A., BARBANTI, M., CARUSO, C., CONIGHI, C., CONTE, R., FAVOINO, B., GALLIANO, L., LUCIANI, G., MAGRI, D., MARTINETTI, M., MATTIUZ, P. L., MENICUCCI, A., MISEFARI, V., MORO, L., PURPURA, M. & SAVI, M. (1989b). The distribution of HLA antigens in Italy. Gene Geography 3, 141-164.
- REYNOLDS, J., WEIR, B. S. & COCKERHAM, C. C. (1983). Estimation of the coancestry coefficient: basis for a short term genetic distance. *Genetics* **105**, 767-779.
- RIORDAN, J. R., ROMMENS, J. M., KEREM, B.-S., ALON, N., ROZMAHEL, R., GRZELCZAK, Z., ZIELENSKI, J., LOK, S., PLAVSIC, N., CHOU, J.-L., DRUMM, M. L., IANUZZI, M. C., COLLINS, F. S. & TSUI, L.-C. (1989). Identification of the cystic fibrosis gene: cloning and characterization of complementary DNA. Science 245, 1066-1073.
- ROMEO, G., BIANCO, M., DEVOTO, M., MENOZZI, P., MASTELLA, G., GIUNTA, A. M., MICALIZZI, C., ANTONELLI, M., BATTISTINI, A., SANTAMARIA, F., CASTELLO, D., MARIANELLI, A., MARCHI, A. G., MANCA,

- A. & MIANO, A. (1985). Incidence in Italy, genetic heterogeneity and segregation analysis of cystic fibrosis. Am. J. Hum. Genet. 37, 338-349.
- ROMMENS, J., KEREM, B. S., GREER, W., CHANG, P., TSUI, L. C. & RAY, P. (1990). Rapid nonradioactive detection of the major cystic fibrosis mutation. Am. J. Hum. Genet. 46, 395–396.
- Saiki, R. H., Walsh, P. S., Levenson, C. H. & Erlich, H. A. (1989). Genetic analysis of amplified DNA with immobilized sequence-specific oligonucleotide probes. *Proc. Natl. Acad. Sci. USA* **87**, 8447–8451.
- SAITOU, N. & NEI, M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol. Biol. Evol. 4, 406-425.
- Sangiuolo, F., Macaratesi, P., Mesoraca, A., et al. (1995). Simultaneous detection of DelF508, G542X, N1303K, G551D and 1717-1G-> A cystic fibrosis alleles by a multiplex DNA enzyme immunoassay. Int. J. Clin. Lab. Res. 25, 142-145.
- Schwarz, M. J., Malone, G. M., Haworth, A., Cheadle, J. P., Meredity, L. A., Gardner, A., Sawyer, H. I., et al. (1995). Cystic fibrosis mutation analysis: report from 22 U.K. regional genetic laboratories. Hum. Mutat. 6, 326-333.
- Watterson, G. A. & Guess, H. A. (1977). Is the most frequent allele the oldest? *Theor. Pop. Biol.* 11, 141–160.
- Welsh, M. J., Tsui, L.-C., Boat, T. F. & Beaudet, A. L. (1995). Cystic fibrosis. In *The metabolic and molecular bases of inherited disease* (ed. C. R. Scriver, A. L. Beaudet, W. S. Sly & D. Valle), pp. 3799-3876. New York: McGraw-Hill Inc. Publ.